

ENZYME PROTEASES USED IN LAUNDRY DETERGENTS ENGINEERING A REVIEW

¹Syed Muhammad Talha Shahid and ¹Kashif Ahmed*

Talha_shahid2011@hotmail.com, Kashif25473@yahoo.com*

¹Department of Chemistry, N.E.D. University of Engineering & Technology, Karachi, Pakistan.

ABSTRACT: *Proteases are fundamental fixings in current laundry detergents. Over the previous years, subtilisin proteases utilized in the detergent industry have been built by guided advancement and sane outline to tailor their properties towards modern requests. This thorough audit talks about late examples of overcoming adversity in subtilisin protease building. Progress in protease building for laundry detergents include concurrent change of warm resistance and movement at low temperatures, a normal methodology to tweak pH profiles, and a general theory for how to increment unbridled action towards the generation of peroxy-carboxylic acids as mellow bleaching specialists. The three protease building effort exhibited give top to bottom investigation of protease properties and have distinguished rule that can be connected to enhance or create chemical variations for mechanical applications past laundry detergents.*

KEY WORDS: *Subtilisin, Thermal Resistance, temperature, Peroxy-carboxylic Acids, Bleaching Agents, PH-Dependent Activity, Fermentation Considerations.*

1. INTRODUCTION

Proteases (proteinases or peptidases) are catalysts that catalyze the hydrolysis of peptide bonds. They are found in all creatures, where they assume a vital part in metabolic and physiological procedures. Substrate unspecific proteases take an interest in protein reusing and assimilation, though succession particular proteases are crucial for zymogene actuation, synergist falls, and other physiological procedures identified with cell survival or passing [1]. Proteases are ordered by their reactant instrument into aspartic, glutamic, serine, cysteine or metalloproteases, by their capacity to divide terminal amino acids as exo or endo peptidases, or by pH conditions for ideal movement (corrosive, nonpartisan or soluble proteases) [2]. Serine proteases are the most copious sort of protease containing a serine as vital synergist amino corrosive buildup. Serine starts a nucleophilic assault on the peptide bond in an electronic situation gave by a neighboring histidine and aspartic corrosive [3]. Further grouping of serine proteases is subject to substrate specificity and basic homology to entrenched proteases [4]. The principle subclasses of serine proteases are subtilisin like, chymotrypsin like, wheat serine carboxypeptidase II-like, prolyligopeptidase like, myxobacter alytic and staphy lococcal proteases [2].

Despite their physiological significance, proteases are of awesome use in modern enzymatic applications, for example, laundry detergents, programmed dishwashing, food added substances, nourishment preparation, cowhide, diagnostics, therapeutics and pharmaceutical industries [2,5]. The significant utilization of proteases in the nourishment business is the upgrade of flavor in dairy, meat, and fish items. In the calfskin and fleece industry, proteases discover their application for splashing, de hairing, and hydrolysis of covering scales on fleece strands [6]. In diverse restorative medications, proteases are utilized as dynamic operators (treatment of osteoarthritis, evacuation of dead tissue, wound mending) [7].

Subtilisin-like proteases are serine proteases created as

additional cell compounds with a sub-atomic weight extending from 18 to 90 kDa and are primarily utilized in industry because of their out-standing properties, for example, high security and wide substrate specificity [5].

2. SUBTILISINS

Subtilisins are characterized by their synergist system as serine proteases. Their amino destructive gathering and three-dimensional structure can be doubtlessly isolated from the other serine proteases, for instance, chymotrypsin, carboxypeptidase and Peptidase A from *Escherichia coli*. The reactant triad of subtilisins comprises of aspartic corrosive, histamine and serine. In spite of the fact that the extent of subtilisins fluctuates from 18 kDa to 90 kDa, all the subtilisins utilized as a part of cleansers have a size of around 27 kDa.

The achievement of subtilisins depends on a few elements, including their high solidness and moderately low substrate specificity highlights regular in extracellular proteases. Their creation as extracellular proteins is obviously an essential component in itself, as this enormously rearranges the detachment of the compound from the biomass and encourages other downstream preparing steps. Another imperative point is the capacity of *Bacillus* strains to discharge catalysts over a brief timeframe into the maturation juices.

Subtilisins are utilized as a part of a wide range of clothing cleansers and in programmed dishwashing cleansers. Their capacity is to corrupt proteinaceous stains [8]; run of the mill stains incorporate blood, milk, egg, grass and sauces. For testing purposes, such stains are economically accessible from test organizations. For programmed dishwashing tests the readiness of stains has been portrayed in extraordinary point of interest. An angle that must be considered while screening applicant chemicals for better execution is that they are not following up on dissolvable substrates in arrangement, but rather on substrates bound to the surface of a strong, water-insoluble substrate.

As opposed to more biochemical situations, where the denaturation of protein substrates ordinarily prompts

enhanced compound action, the denaturation of proteinaceous stains by maturing, warming and oxidization makes them less available to chemical debasement. The impact of oxygen dye on warmth denatured blood or drain stains is an incredible case: the vicinity of oxygen fade changes a generally simple protease focus into a to a great degree troublesome one. Along these lines, test results depend basically on the sort of stain, the piece of the cleanser and the nature and status of the materials utilized as a part of the washing test as filler material (weight). The same remains constant for programmed dishwashing cleansers, where the nature, synthesis and measure of weight stain are vital perspectives in the assessment of compound execution.

3. THE IDENTIFICATION & OPTIMIZATION OF DETERGENT PROTEASE

At present, fewer than 15 distinctive protein atoms are utilized as a part of detergents around the world. These chemicals start from *B. amyloliquefaciens*, *B. licheniformis*, *Bacillus clausii*, *Bacillus lentus*, *Bacillus alkaloophilus*, and *Bacillus halodurans* [2]. Some of these species assignments have as of late been changed; for instance, Savinase1 and Esper-asee1 were doled out for quite a while to *Bacillus subtilis* or *B. lentus* [5, 6].

3.1. Protein Engineering Methods

Since protein designing with subtilisins, all the amino corrosive positions have been altered either by site coordinated mutagenesis in light of balanced outline or, later, by different routines for irregular mutagenesis. A large portion of this work is distributed in licenses and not in the investigative writing. Hydrogen peroxide and peroxy acids are run of the mill fading operators created in the cleaning procedure of fade containing items. The oxidation of certain methionine deposits to sulfoxides was known for over 10 years before the first ways to deal with site-coordinated mutagenesis were figured it out. In 1997 the first proteases changed along these lines for hydrogen peroxide steadiness were advertised, despite the fact that the execution of these variations did not satisfy their guarantee. By 1999 substitutions at about each position in the adult 275 amino corrosive BPN0 subtilisin (*Bacillus Protease Novo* sort, subtilisin from *B. amyloliquefaciens*) had been asserted in licenses. The BPN0 subtilisin is for the most part thought to be the lead atom for subtilisin alterations, and transformations in other subtilisins frequently allude to the homologous position in this lead particle. There are some amazing general surveys on the protein building of subtilisin, and in addition articles on more particular detergent applications.

Since, a few quality rearranging methodologies have been performed with subtilisins. Fascinating results from the rearranging of 26 protease qualities have been portrayed for properties, for example, action in natural solvents, temperature soundness, and movement at high or low pH [10, 12]. Little, then again, has been distributed on stain evacuation. Attributable to the substantial number of variation particles generated by rearranging and other

arbitrary strategies, screening routines with high-throughput and expanded significance have must be produced. Shockingly, these techniques are still not by any means tasteful, which may clarify why no exceptional new subtilisin variation made by one of the quality rearranging advancements is yet present in detergents.

Phage show, as a result of the nearby connection between the polypeptide and its encoding quality, is thought to be an amazing system for the choice of chemicals with sought properties. Legendre et al. [13] utilized subtilisin 309 to represent the capability of this system, changing its substrate specificity concerning the amino destructive at the P4 site of the substrate. Soumillion and Fastrez [14] give further specimens of phage showcase applications with subtilisins. As all transformations influencing the specificity of subtilisins may likewise impact the autoproteolytic preparing of the proenzyme to the adult shape, the designing of the genius area or its uncoupling from this biosynthesis step has ended up applicable [15,16]. In this way, no additional data on the, Accomplishment of such methodologies has turned out to be openly accessible.

3.2. Proteases As Additives In Laundry Detergent Applications

Proteases as trypsin and chymotrypsin were presented surprisingly as a dynamic fixing in laundry detergent prevent gentlemen for debasement of proteinaceous stains in 1995 by the German scientific expert Otto Ro'hm [8]. The main business cleanser containing bacterial proteases was delivered by Gebruder Schnyder in 1996. By 1996, around 70% of the substantial obligation clothing detergent in Europe contained compounds. In the most recent 50 years, proteases and different proteins in clothing detergent changed from being minor added substances to key fixings. The determination and assessment of proteases to be utilized as a part of detergent depends on essential parameters characterized by clothing cleanser makers. A cleanser protease needs productive washing execution at expansive soluble pH and over an extensive variety of temperatures (from low temperatures for engineered strands, to high temperatures for cotton). The execution of a decent cleanser protease is characterized by various parameters, for example, proteinaceous stain corruption, similarity with other cleanser segments (e.g. nonionic and anionic surfactants, complexion specialists, per-vapor, and different chemicals), solidness in the vicinity of oxidizing operators as fade, and time span of usability in cleanser plans. The main compound suppliers and cleanser makers are effectively seeking after the advancement of new chemical exercises that address shopper requirements for enhanced cleaning, fabric care and antimicrobial properties. Subsequently, look into on proteases has concentrated on revelation and building catalysts that are more hearty as for pH, temperature, dependability and substrate specificity By utilizing methods of protein designing and objective outline.

In this survey we will concentrate on three critical properties for the use of subtilisin proteases in the clothing cleanser industry that have been handled by protein building; action and warm resistance of *Bacillus gibsonii* basic protease (BgAP) was at the same time expanded [9], unbridled action of subtilisin Carlsberg was expanded towards peroxycarboxylic acids creation [10] and the pH movement profile of BgAP was moved towards higher action at lower (pH range 8.5-1).

4. PRODUCTION ASPECTS

4.1. Production Strains

All major subtilisins for detergents are delivered in *Bacillus*, in light of the fact that these species can discharge a lot of extracellular catalyts [23]. The control instruments included in the generation of proteases in *Bacillus* are to a great degree mind boggling and still not completely understood. A sample is the two-part administrative framework that goes about as a majority detecting instrument in *B. subtilis* and which has been found to control the declaration of the soluble protease [24]. This administrative framework is encoded on the chromosome and on endogenous plasmids.

Modern strain change projects utilizing traditional microbiological techniques have been completed over numerous years and have brought about the improvement of a few profoundly beneficial strains. These have regularly been utilized as hosts for the declaration of recombinant qualities; on the other hand, these modern creation strains have much of the time been depicted as impervious to change. As plasmid based creation strains regularly show insecurity master blems, it is presently standard practice to produce strains in which the recombinant quality is coordinated into the chromosome in numerous duplicates [25, 26].

4.2. Hot And Cold, The Temperature Challenge Of Modern Detergent Proteases

These days' patterns in vitality effectiveness bring the mindfulness up in the public eye for washing at low temperature. Outlining reasonable laundry detergents with elite at low temperatures requires the improvement of catalyts with high efficiency at wide temperature extend particularly at temperatures <208C [12]. Proteases adjusted to low temperatures can be detached from normally happening psychrophilic microorganisms, showing high proteolytic movement (158C) [13, 15].

Tragically, such compounds for the most part don't meet modern prerequisites because of natural low solidness at temperatures above 208C and low item yields in huge scale creation [5, 12, 16]. Then again, subtilisins disconnected from mesophilic creatures display in the meantime higher reactant efficiency and temperature security at temperatures from 308C to 458C. So as to adjust mesophilic subtilisins to the present pattern of washing at low temperature (208C) coordinated advancement and judicious configuration methodologies have been utilized. One coordinated development crusade was performed with *Bacillus sphaericus* subtilisin (SSII)

utilizing arbitrary mutagenesis took after by recombination of enhanced variations. A striking increment in the turnover number (kcat at 108C expanded 6.6 overlap) and expanded synergist efficiency (9.6 fold higher than wild sort) was accomplished [17].

In another methodology a chimeric protein was created by supplanting the exceedingly flexible 12 amino acid. The coming about half and half compound demonstrated a higher specific movement for engineered substrates and a widened substrate profile at room temperature [18]. Security of subtilisins is a property which has been concentrated broadly by arbitrary mutagenesis and normal methodologies. In an escalated investigation, amino corrosive substitutions and their belongings in more than half of the 275 amino acids of subtilisin [19]. Research on subtilisin adjustment concentrated on calcium subordinate and autonomous solidness, and also adjustment by the presentation of disulfide bonds [19, 25]. Notwithstanding, a general instrument portraying the security of proteases must be illustrated. In many proteins, intermolecular communications, for example, salt scaffolds are fundamental for warm steadiness. It has been demonstrated that warm dependability was significantly diminished by evacuation of salt scaffold systems in aqualisin I, a thermo stable subtilisin protease [26, 27]. In another methodology, the clearly inverse properties of high warm strength in blend with expanded action at low temperatures were examined in *B. gibsonii* soluble prtease (BgAP)[9].

4.3. Engineering Of pH-Dependent Activity

The achievement of subtilisins for their application in detergents depends, as talked about some time recently, on a few components, for example, similarity with the detergent network, wide substrate specificity, warm resistance, action at temperatures from 208C to 608C, and in addition high action in a wide soluble pH range. Industrially pertinent compounds for application in detergent industry are subtilisin proteases started from *Bacillus* sp. including the broadly utilized proteases of *Bacillus amyloliquefa-ciens* (subtilisin BPN') [34, 35], *B. gibsonii* (BgAP) [9, 11], *B. lentus* (subtilisin BL) [36], and *Bacillus licheniformis* (subtilisin Carlsberg) [37,38]. The identification of new basic proteases is a progressing test [39]. Action at high basic pH is an essential for protease application in detergent plans. So far the pH subordinate action of subtilisins or hydrolysis as a rule is not totally caught on. A few endeavors to balance pH subordinate action concentrated on the designing of charge appropriation, since the pH subordinate movement is essentially dictated by the pKa estimations of the dynamic site deposits. Thusly "vast" pH-movement profile movements are regularly identified with changes situated in close vicinity to the dynamic site which sadly build the opportunity to get chemical variations with lessened or no movement. Amino corrosive substitutions removed from the dynamic site, for example, surface uncovered buildups, generally bring about catalyts variations keeping

up the wild sort action and displaying a "little" move in the pH profile [40]. The influence of protease surface charges on the pH subordinate subtilisin action was exhibited as of now by amino corrosive substitutions in 1997 [41]. As of late another amino corrosive determination methodology to build the pH subordinate action of the *B. gibsonii* antacid protease (BgAP) (pH ideal 11), taking after the deamidation standard, was accounted for. The posttranslational autocatalytic deamidation procedure changes over asparagine and glutamine buildups into adversely charged aspartic corrosive and glutamic corrosive, separately.

This procedure changes the net charge of the proteins and consequently their pH subordinate action. This deamidation rule was imitated by site coordinated mutagenesis (Gln to Glu and Asn to Asp). Three criteria for amino corrosive choice were defined by (1) amino acids of the deamidation sort (Asn or Gln), (2) non-monitored buildups and (3) surface uncovered deposits neighbored by glycine. The protease BgAP comprises of 270 amino acids, wherein 113 amino acids are surface uncovered and 31 of the surface uncovered buildups are Asn or Gln deposits. For assessment 18 singular substitutions (11 Asn and 7 Gln) fulfilling one, two or every one of the three criteria were chosen and created. Site-coordinated mutagenesis taking after the deamidation guideline in five (Asn97, Asn253, Gln37, Gln200, and Gln256) out of eight (Asn97, Asn154, Asn250, Asn253, Gln37, Gln107, Gln200, and Gln256) amino acids meeting every one of the three criteria brought about expanded proteolytic action at pH 8.6. Variations Asn253Asp and Gln256Glu and the consolidated variation Asn253Asp/Gln256Glu demonstrated a pH ideal at 10 (wild sort pH ideal 11). The joined variation Asn253Asp/Gln256Glu indicated 2 fold increment in action at pH 8.5 contrasted with the wild sort [11].

The criteria introduced of the deamidation rule are in principle free of the protein/chemical class together with the all inclusive statement in regards to the impact of surface charge changes on pH subordinate catalyst movement, it is likely that properties of compounds from diverse classes can be modified utilizing the deamidation approach.

4.4. Granulation Process

Subtilisin arrangements are promoted either as a settled chemical arrangement or as embodied and covered granulates. The fluid arrangements typically have a diminished water content and contain huge measures of 1, 2-propane diol. The first grind sort (prill), which depended on a blend of chemical and polyethylene glycol, has now for all intents and purposes vanished from the business sector. Granulation procedures incorporate the utilization of expulsion, high shear blending and fluidized

beds [33]. The gear utilized for granulation additionally decides the kind of crush created in the process regarding shape, substance arrangement and structure. In all cases, one or a few covering layers guarantee that grinds have low cleaning properties. This prerequisite results from the acknowledgment in 1995 that compound dust, produced amid the detergent assembling procedure, can prompt the sensitization of uncovered specialists. Granulation innovation was created to maintain a strategic distance from the arrival of this protein containing dust. This innovation has now been further enhanced and extra strides in the generation process (e.g. embodiment and ventilation) have been acquainted with wipe out the issue. Preparing and different control instruments, as a major aspect of word related wellbeing projects, have likewise been set up in the Detergent Industry.

4.5. Catalytic Promiscuity in Subtilisin Proteases:

Switching Proteolysis Towards Perhydrolysis

One of the key segments of present day powder laundry detergent is the vicinity of oxygen based dyeing specialists. Regularly, hydrogen peroxide is utilized as an oxidative specialists delivered by unconstrained deterioration of per borate and per carbonate consolidated with tetra-acetyl ethylenediamin or no nanoyloxy benzene sulphonate [42]. The convergences of artificially created hydrogen peroxide are normally high and hurtful for material and surfaces and in addition information examination the favored ester substrate for subtilisin Carlsberg variations is methyl-butyrate (k_{cat} qualities are significantly expanded contrasted with the subtilisin Carlsberg wild sort) while the KM qualities are decreasing with development in the substrate chain length. The last, proposes the favoring of hydrophobic coordinated efforts between stores in the S1 tying pocket and the ester substrates. The amino corrosive substitutions Gly165Leu, Gly165Ile, and Gly165-Tyr brought about a size lessened S1 tying pocket and hence influenced the coupling introduction of the ester substrates. Substitutions of Gly by more massive amino acids (Leu, Ile, and Tyr) came about just in the beneficial tying method of the substrate contrasted with the subtilisin Carlsberg while sort in which non gainful and profitable mode have been watched. The beneficial tying mode (ester gathering of the substrate is found near the synergist buildup Ser220) saw in every one of the three variations gives a first speculation to clarify expanded level of perhydrolysis (Fig.1) [10].

This work proposed that amino acids at position Gly165 are critical for substrate specificity in the peroxycarboxylic corrosive generation. Three substitutions at position Gly165 changed over the protease subtilisin Carlsberg into a perhydrolyase with practically identical total exercises of regular perhydrolysis.

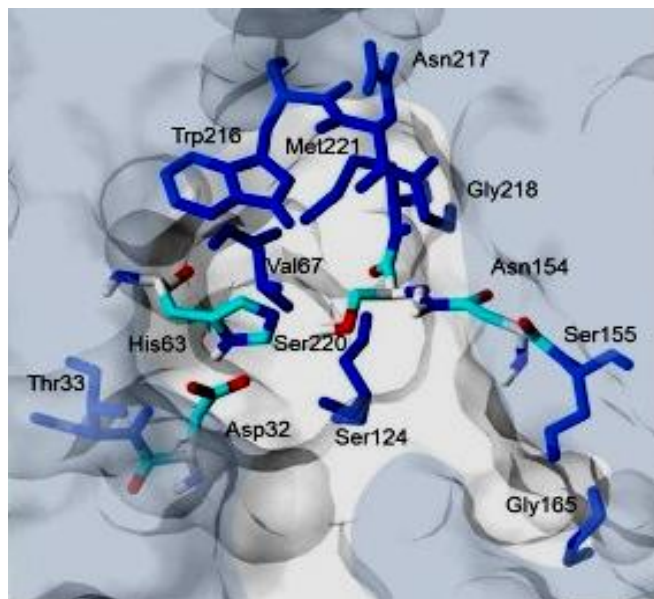


Fig. 1 Expanded level of perhydrolysis

Computational investigation recommends that diminishment in the measure of the coupling S1 pocket is a key for advancing perhydrolysis. This speculation can be further connected to enzymatic in situ creation of peroxy-carboxylic corrosive and can widen its application to beauty care products, cleanser, mash, and calfskin commercial enterprise.

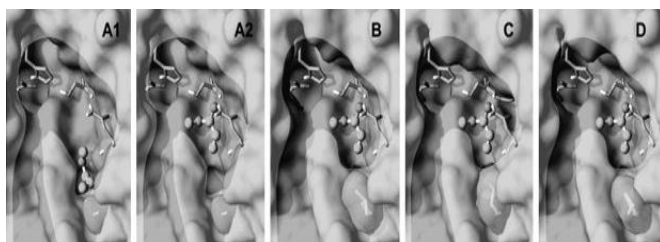


Fig. 2 Hypothetical effect of mutations at position 165 in the S1 pocket for the catalysis of methylesters (methyl propionate).

Fig. 2 shows the hypothetical effect of mutations at position 165 in the S1 pocket for the catalysis of methylesters (methyl propionate). Docking studies revealed that the modulation of the S1 pocket can change the configurations in which the methyl ester binds to the protease active site. In the wild type, there are two possible modes of the substrate (A), of which only one is in the productive conformation (A2). When Gly165 is substituted by bulkier Ile, Leu or Tyr residues (B, C, D), the non-productive binding mode does not occur, possibly increasing the reaction rate [10].

4.6. FERMENTATION CONSIDERATIONS

The fermentation of subtilisins is liable to a substantial number of variables. The parameters range from the media arrangement, Ph and oxygen-exchange rate to the distinctive *Bacillus* species utilized as hosts for recombinant creation. Mechanical creation procedures are ordinarily keep running as large-scale, sustained clump maturations at high cell thickness. Persistent maturations

are utilized to dissect basic creation parameters, however are not utilized for generation [30]. The media utilized as a part of mechanical maturations have, most importantly, to satisfy monetary necessities; consequently, these media are regularly in view of unpredictable, economical nitrogen sources. The arrangement of maturation media and the points of interest of the aging procedures and yields are regularly considered organization insider facts and in this manner no solid data is accessible in people in general area. The general standards on aging and downstream handling have been distributed, however [31]. As of late, distributions on the generation of modern *Bacillus* strains have depicted yields in the scope of 20–25 g/L protein in maturation stock [32].

4.7. Formulation In Detergents

Granulated proteins place couple of limitations on the plan of powder detergent and tablets. Increased contact with dying parts, experienced for instance in the squeezing of tablets, requires more elevated amounts of capacity soundness. This thus has advanced the improvement and utilization of oxidation stable catalyst variations. The adjustment of proteases in fluid arrangements is still a field for exploration [34, 35]. The real issue in fluid situations is auto proteolysis. Some broad standards in defining fluid detergent incorporate the diminishment of the free water focus and the utilization of reversible inhibitors like borate or phenyl boronic corrosive subordinants. What's more, the piece and nature of the surfactants in the fluid cleanser extraordinarily impact the capacity strength of the compound. Fluid detergent must be planned around the requirements of the chemicals they contain, streamlining approaches to settle and restrain them reversibly.

5. CONCLUSION

Subtilisins are imperative mechanical chemicals and crucial added substances in cutting edge removing so as to laundry detergents to support washing execution efficiently proteinaceous stains. Reported examinations concentrate on the designing of the subtilisin BgAP towards expanded action at low temperatures (158C) and concurrent change of warm resistance; two opposite properties requiring solid sub atomic cooperation and legibility in one compound. Furthermore, a general relevant objective protein building technique to tailor subtilisin surface charges with a specific end goal to change pH subordinate action was exhibited. At long last, the substitution of one essential amino corrosive position in the S1 specificity pocket prompted a decreased pocket size, which turned subtilisin.

ACKNOWLEDGEMENT

Authors want to thank the professors of the Departments, Unilever Team and deep Cooperation by FutehAlly Chemicals Company for their help and careful proof reading of the manuscript.

REFERENCES

- [1] Siezen RJ, Leunissen JAM: Subtilases: the superfamily of subtilisin likes serine proteases. *Protein Sci* (1997), 6:501-(523).
- [2] Egmont MR: submission of proteases in detergents. In *Enzymes in Detergency*. Edited by Van Ee J, Misset O, Baas EJ. New York: Marcel Dekker Inc., Surfactant Science Series (1997), 69: 61- (74).
- [3] Enzymes in household detergents: In *Enzymes in Industry*. Edited by Aehle W. Ullmann's Encyclopedia of Industrial Chemistry Chapter 5.2.1. Weinheim: Wiley VCh Verlag (2004):155-(180).
- [4] Detailed overview on enzyme effects in laundry detergents in relation to detergent composition and geographic differences in washing conditions, including the test materials used for enzyme evaluation.
- [5] Enzymes in automatic dishwashing: In *Enzymes in Industry*. Edited by Aehle W. Ullmann's Encyclopedia of Industrial Chemistry Chapter 5.2.2. Weinheim: Wiley VCh Verlag (2004):180-(194).
- [6] Outtrup H, Jorgensen ST: The importance of Bacillus species in the production of industrial enzymes. In *Applications and Systematics of Bacillus and Relatives*. Edited by Berkeley R, Heyndrickx M, Logan N, De Vos P. Oxford, UK: Blackwell Science Ltd. (2002): 206-(218).
- [7] Review on the relevance of Bacillus for industrial enzyme production from the position of the market leader.
- [8] Ito S, Kobayashi T, Ara K, Ozaki K, Kawai S, Hatada Y: Alkaline detergent enzymes from alkaliphiles: enzymatic properties, genetics, and structures. *Extremophiles* (1998), 2:185-(190).
- [9] Estell DA, Graycar TP, Wells JA: Engineering an enzyme by site directed mutagenesis to be resistant to chemical oxidation. *J Biol Chem* (1985), 260:6518-(6521).
- [10] Bryan PN: Protein engineering of subtilisin. *Biochim Biophys Acta* (2000), 1543:223-238. (334) Protein technologies and commercial enzymes Comprehensive review of the protein engineering work on subtilisin published in the scientific literature.
- [11] Bott R: Development of new proteases for detergents. In *Enzymes in Detergency*. Edited by Van Ee J, Misset O, Baas EJ. New York: Marcel Dekker Inc., Surfactant Science Series (1997), 69: 75-(91).
- [12] Ness JE, Welch M, Giver L, Bueno M, Cherry JR, Borchert TV, and Stemmer WPC, Minshull J: DNA shuffling of subgenomic sequences of subtilisin. *Nat Biotechnol* (1999), 17:893-(896).
- [13] Ness JE, Kim S, Gottman A, Pak R, Krebber A, Borchert TV, Govindajaran S, Mundorff EC, Minshull J: Synthetic shuffling expands functional protein diversity by allowing amino acids to recombine independently. *Nat Biotechnol* (2002), 20:1251-(1255).
- [14] Wintrode PL, Miyazaki K, Arnold FH: Cold adaptation of a mesophilic subtilisin like protease laboratory evolution. *J Biol Chem* (2000), 275:31635-(31640).
- [15] Legendre D, Laraki N, Gra'slund T, Bjørnvad ME, Bouchet M, Nygren P-A, Borchert TV, Fastrez J: Display of active subtilisin 309 on phage: analysis of parameters influencing the selection of subtilisin variants with changed substrate specificity from libraries using phosphonylating inhibitors. *J Mol Biol* (2000), 296:87-(102).
- [16] Soumillion P, Fastrez J: Investigation of phage display for the directed evolution of enzymes. In *Directed Molecular Evolution of Proteins*. Edited by Brakmann S, Johnsson K. Weinheim: Wiley VCH Verlag; (2002): 79-(110).
- [17] Takagi H, Takahashi M: A new approach for alteration of protease functions: pro-sequence engineering. *Appl Microbiol Biotechnol* (2003), 63:1-(9).
- [18] Almog O, Gallagher T, Tordova M, Hoskins J, Bryan P, and Gilliland GL: Crystal structure of calcium-independent subtilisin BPN⁰ with restored thermal stability folded without the prodomain. *Proteins* (1998), 31:21-(32).
- [19] Cherry JR, Fidantsef AL: Directed evolution of industrial enzymes: an update. *Curr Opin Biotechnol* (2003), 14:438-(443). Most recent review on the directed evolution of enzymes, including a section on proteases.
- [20] Saeki K, Hitomi J, Okuda M, Hatada Y, Kageyama Y, Takaiwa M, Kubota H, Hagihara H, Kobayashi T, Kawai S, Ito S: A novel species of alkaliphilic Bacillus that produces an oxidatively stable alkaline serine protease. *Extremophiles* (2002), 6:65-(72).
- [21] Saeki K, Hitomi J, Okuda M, Hatada Y, Kobayashi T, Ito S, Takami H, Horikoshi K: Novel oxidatively stable subtilisin-like serine proteases from alkaliphilic Bacillus spp.: enzymatic properties, sequences and evolutionary relationships. *Biochem Biophys Res Commun* (2000), 279:313-(319).
- [22] Gupta R, Beg QK, and Lorenz P: Bacterial alkaline proteases: molecular approaches and industrial application. *Appl Microbiol Biotechnol* (2002), 59:15-(32).
- [23] Pogson M, Georgiou G, Iverson BL. Engineering next generation proteases. *Curr Opin Biotechnol* (2009); 20:390-(7).
- [24] Gupta R, Beg QK, Khan S, Chauhan B. An overview on fermentation, down-stream processing and properties of microbial alkaline proteases. *Appl Microbiol Biotechnol* (2002); 60:381-(95).
- [25] Wells JA, Estell DA. Subtilisin an enzyme designed

- to be engineered. *Trends Biochem Sci* (1988); 13:291–(7).
- [27] Morihara K. Comparative specificity of microbial proteinases. *Adv Enzymol Relat Areas Mol Biol* (2006); 41:179–(243).
- [28] Maurer KH. Detergent proteases. *Curr Opin Biotechnol* (2004); 15:330–(4).
- [29] Varela H, Ferrari MD, Belobrajdic L, Weyrauch R, Loperena L. Short communication: effect of medium composition on the production by a new *Bacillus subtilis* isolate of protease with promising unhairing activity. *World J Microbiol Biotechnol* (1996); 12:643–(5).
- [30] Mekkes JR, Le Poole IC, Das PK, Bos JD, Westerhof W. Efficient debridement of necrotic wounds using proteolytic enzymes derived from Antarctic krill: a double-blind, placebocontrolled study in a standardized animal wound model. *Wound Repair Regen* (1998); 6:50–(7).
- [31] Herbots I, Kottwitz B, Reilly PJ, Antrim RL, Burrows H, Lentin g HBM, et al. *Enzymes*,
- [32] Non-food application, in Ullmann's encyclopedia of industrial chemistry, Ullmann's encyclopedia of industrial chemistry. Wiley-VCH Verlag GmbH & Co. KGaA; (2000).
- [33] Martinez R, Jakob F, Tu R, Siegert P, Maurer KH, Schwaneberg U. Increasing activity and thermal resistance of *Bacillus gibsonii* alkaline protease (BgAP) by directed evolution. *Biotechnol Bioeng* (2013); 110:711–(20).
- [34] Despotovic D, Vojcic L, Blanusa M, Maurer KH, Zacharias M, Bocola M, et al. Redirecting catalysis from proteolysis to perhydrolysis in subtilisin Carlsberg. *J Biotechnol* (2013); 167:279–(86).
- [35] Jakob F, Martinez R, Mandawe J, Hellmuth H, Siegert P, Maurer KH, et al. Surface charge engineering of a *Bacillus gibsonii* subtilisin protease. *Appl Microbiol Biotechnol* (2013);97:6793–(802).
- [36] Gerday C, Aittaleb M, Bentahir M, Chessa JP, Claverie P, Collins T, et al. Coldadapted enzymes: from fundamentals to biotechnology. *Trends Biotechnol* (2000); 18:103–(7).
- [37] Arnorsdottir J, Kristjansson MM, Ficner R. Crystal structure of a subtilisin like serine proteinase from a psychrotrophic *Vibrio* species reveals structural aspects of cold adaptation. *FEBS J* (2005); 272:832–(45).
- [38] Cavicchioli R, Siddiqui KS, Andrews D, Sowers KR. Low temperature extremo-philic and their applications. *Curr Opin Biotechnol* (2002); 13:253–(61).
- [39] Davail S, Feller G, Narinx E, Gerday C. Cold adaptation of proteins. Purification, characterization, and sequence of the heat-labile subtilisin from the Antarctic psychrophile *Bacillus TA41*. *J Biol Chem* (1994); 269:17448–(53).
- [40] Saeki K, Ozaki K, Kobayashi T, Ito S. Detergent alkaline proteases: enzymatic properties, genes, and crystal structures. *J Biosci Bioeng* (2007); 103:501–(8).
- [41] Wintrode PL, Miyazaki K, Arnold FH. Cold adaptation of a mesophilic subtilisin like protease by laboratory evolution. *J Biol Chem* (2000); 275:31635–(40).
- [42] Tindbaek N, Svendsen A, Oestergaard PR, Draborg H. Engineering a substrate-specific cold adapted subtilisin. *PEDS* (2004);17:149–(56).
- [43] Bryan PN. Protein engineering of subtilisin. *Biochim Biophys Acta* (2000); 1543:203–(22).
- [44] Almog O, Gallagher T, Tordova M, Hoskins J, Bryan P, Gilliland GL. Crystal structure of calcium independent subtilisin BPN' with restored thermal stability folded without the prodomain. *Proteins Struct Funct Bioinf* (1998); 31:21–(32).
- [45] Braxton S, Wells JA. Incorporation of a stabilizing Ca²⁺-binding loop into subtilisin Bpn'. *Biochemistry* (1992); 31:7796–(801).
- [46] Bryan P, Alexander P, Strausberg S, Schwarz F, Lan W, Gilliland G, et al. Energetics of folding subtilisin Bpn'. *Biochemistry* (1992); 31:4937–(45).
- [47] Mitchinson C, Wells JA. Protein engineering of disulfide bonds in subtilisin Bpn'. *Biochemistry* (1989); 28:4807–(15).
- [48] Pantoliano MW, Whitlow M, Wood JF, Rollence ML, Finzel BC, Gilliland GL, et al. The engineering of binding-affinity at metal ion binding-sites for the stabilization of proteins subtilisin as a test case. *Biochemistry* (1988); 27:8311–(7).
- [49] Takagi H, Takahashi T, Momose H, Inouye M, Maeda Y, Matsuzawa H, et al. Enhancement of the thermostability of subtilisin E by introduction of a disulfide.